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## Food Microbiology Reference Tool

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## **Targets of Irradiation – What are they and how do we kill or inactivate them?**

Understanding the differences between the agents of illness and spoilage can help to understand how or if irradiation can kill or inactivate them.

Most spoilage or pathogenic (harmful) agents in foods are microorganisms, which are commonly considered to be bacteria, yeasts and molds. Other harmful agents include insect pests, parasites, prions and viruses. The FDA has an online reference book for pathogenic microorganisms and parasites at: <http://vm.cfsan.fda.gov/~mow/intro.html>

Bacteria – Small single cells of various shapes. Bacteria replicate the same way (replicate the cell's genetic matter and then divide the cell in two). Consequently, if irradiation harms the cell structure or genetic material, the process can kill the bacterial cell. Bacteria are also sensitive to their environment, so chemical changes caused by irradiation can damage or kill them. Some bacteria can form tough dormant spores in response to hostile environments and these spores are very difficult to kill with irradiation or other means. Bacteria are classified by their acceptance of Gram stain. Gram-positive bacteria are often slightly more likely to tolerate irradiation, but you can't definitely rely on this to tell you the bacteria's response to radiation.

Fungi - Fungi include molds and yeasts. Usually more difficult to kill than bacteria because of their shape, growth characteristics and adaptability.

Molds – Fast growing multicellular fungi of various shapes, but mostly stringy. They multiply in many different ways. Under a microscope, molds look like various forms of sea coral. They are often stringy and if a piece is broken by irradiation, the broken piece is often still capable of growth. This is one reason they are difficult to kill. Molds are the most tolerant microorganism to changes in pH, water content and oxygen, so the environmental changes caused by irradiation may not affect them as much.

Yeasts – Single cell fungi, but some have stringy parts. The cells are larger than bacteria and of various shapes. Can replicate in different ways, making them somewhat more difficult to kill than bacteria. Yeasts that survive irradiation can sometimes grow very quickly, resulting in quick spoilage of some irradiated foods.

Insect pests – I use this term to include insects, spiders and the other small creatures that have various life stages and are sometimes living in or hitchhiking in food. They can carry disease, but often are not actually harmful if eaten (unless you see them!). The problem is they can be harmful to agriculture and spoil food with their excrement, webs and living structures. Insect pests damage food quality and are generally distasteful. Insect pests vary in their sensitivity to radiation depending on the life stage. Rapid growth stages are usually less tolerant to radiation. So, usually, eggs are more sensitive than are larvae; following larvae, the pupae (cocoons) are more sensitive and finally adults are very difficult to kill, but easy to sterilize.

Parasites – Small animals that live their lives in other animal hosts. They are much larger than bacteria or fungi (if you apply a stain to the food you can often see them, even without magnification), and have several life stages (egg, larvae, adult). They may develop in foods, but unlike bacteria, parasites do not proliferate in foods. They are often easier to kill because they are sensitive to their environment and if one life stage is damaged, it might not be able to proceed to the next life stage. It is less difficult to characterize the radiation tolerance of parasites, in part because much less research has been done on them.

Prions - extremely small protein-like fractions that, like viruses, are not alive. Prions are considered to be the infective agent for Transmissible Spongiform Encephalopathies (TSE) such as Scrapie (a disease of sheep) and 'Mad Cow' disease, the Bovine form of the disease, shortformed as BSE. I have seen nothing that would indicate prions could be inactivated by irradiation at any reasonable dose.

Viruses – protein-protected DNA or RNA, these agents use their hosts's biological components to create havoc, but viruses are not actually alive. Therefore we don't kill viruses, we inactivate them. Since protein is not much affected by irradiation (and the proteins around viruses are tough), irradiation has to directly hit the DNA or RNA and break it to inactivate a virus. This is one reason why inactivating viruses usually requires a higher dose than killing bacteria. Viruses vary in their sensitivity to environment so chemical change may or may not inactivate them. They are usually quite heat sensitive. Although high coliform counts are usually a good indicator of bacterial contamination, it isn't true with viruses. So, shellfish taken from water with low coliform counts can still be quite contaminated with viruses.

## Common Foodborne Spoilage Bacteria - Quick Notes

*Coliform* – Gram-negative. A collective name for several members of the Enterobacteraceae family that includes some pathogens and some indicators of spoilage. Generally a coliform measurement is an indicator of fecal contamination, and not necessarily an indication of the presence of pathogens. Coliforms are everywhere, including probably all foods. Therefore, we shouldn't confuse a coliform count with the presence (or not) of *E. coli*, which is part of the same family but a different species.

*Lactobacillus* – Gram-positive. Similar to *Leuconostoc*. Spoilage organisms found everywhere but mostly found in plant products and dairy products, and also can spoil processed meat products. Acid and cold tolerant.

*Leuconostoc* – a Gram-positive spoilage bacterium most commonly found in plant products. Also implicated in spoilage of fresh milk and dairy, fresh refrigerated dough products and raw and processed meats (will grow in vac-packed and MAP meats and dough even at refrigeration temps). Similar to *Lactobacillus*, this bacteria is a lactic acid generator. This species is commonly used as starter cultures in dairy processing, and also some fermented beverages because it generates ethanol.

*Pseudomonas* – Gram-negative. By far the most important refrigerated food spoilage organisms. Also commonly implicated in contamination of cosmetics where they can cause skin and eye illnesses.

## Common Foodborne Pathogenic Bacteria - Quick Notes

*Bacillus species* - Gram-positive sporeformers. The need for and difficulty of killing *Bacillus* species spores are one of the reasons that dry foods and ingredients have to be irradiated at higher doses.

*Bacillus cereus* –pathogen of rice, spices, flour, dry soup mixes. This bacteria usually causes illness by producing a toxin that is consumed in reheated or inadequately heated foods. For example, the spore found in rice will usually not cause illness if the rice is eaten immediately after cooking. But if you take the cooked rice and stuff a turkey, or leave the rice at warm temperatures, illness can easily result.

*Bacillus subtilis* – Gram positive sporeformer. Originates in soil; causes spoilage of organism of cereals and flour and herbs and spices. *B. subtilis* in spices and soup mixes has been implicated in foodborne illness (once the spores are activated by heat and being placed in moist foods, they germinate, a toxin is produced which cause illness when the leftovers are eaten).

*Campylobacter* – Gram-negative. (Pronounced Camp - e - lobacter). A pathogen of meat origin and found on most meat carcasses of all species. Even though it is quite heat and freezing sensitive it is considered to cause more illness than any other pathogen, but it is either reported less or lab tests were insufficient to correctly label it. Although the best known pathogen in this species is *Campylobacter jejuni*, several other *Campylobacter*s also cause illness. *Campylobacteriosis* is considered a mild illness, but it has been found to sometimes cause Guillume Barre syndrome a very serious and long term neurological disability.

*Clostridium* – Gram-positive sporeformers, heat tolerant, grows only in low oxygen situations, but low oxygen situations can be found in surprising places, such as under mold or deep in muscle tissue. Species includes several pathogens. Of soil and water origin they are also important pathogens of meat. Illness is caused by the toxins produced by *Clostridium* species.

*Clostridium botulinum* – Forms heat and radiation resistant spore; vegetative cells produce toxin. Probably the most difficult bacteria to kill and, if the food allows the growth of this organism, it is the most important organism to kill. Causes one of the most serious forms of foodborne illness -botulism.

*Clostridium perfringens* – Heat resistant sporeformer; vegetative cells produce toxin. Found in soils, foods are also commonly contaminated with this microorganism by humans. Found in spices, meats and poultry and meat gravies. Causes diarrhea with severe pain.

*E. coli* – Gram negative. (The E. stands for *Escherichia* which is hard to pronounce so no one does.) These bacteria cause illness by producing toxins, but unlike *Bacillus*, *Clostridium* or *Staph* species, the toxin is not formed in the food. *E. coli* toxins are formed inside the person who ate the bacteria. Some cause severe gastro-intestinal illness, and one serotype O157:H7 causes kidney damage, organ breakdown and sometimes death.

*Listeria* – Gram positive. One of the more common food pathogens, *Listeria monocytogenes* is increasingly implicated in illness from refrigerated foods. *Listeria* is one of the more radiation resistant food-borne pathogens, especially at lower temperatures. Although healthy adults may not become ill from *Listeria*, the fatality rate for persons who become ill from *Listeria* is high, (about 34% from illness studied in the 1980's). Listeriosis of pregnant women can cause fetal death and stillborne births.

*Salmonella* – Gram-negative. Several *Salmonella* serovars cause illness. Very widely distributed, especially in animals or animal products (such as milk and eggs). Ingestion of *Salmonella* causes serious illness and can cause particularly disabling forms of arthritis. Salmonellosis is the most common reported foodborne illness in the United States.

*Shigella* – Gram-negative. Three *Shigella* species are foodborne, but essentially they all cause illness from human contamination of food. Similarly to *E. coli*, *Shigella* forms a toxin inside whoever consumes it. Since it causes a serious illness, it is often reported to health authorities. It is the third most reported foodborne illness.

*Staphylococcus aureus* – Gram positive, toxin producer. Several *Staph* species also cause foodborne illness, but this one is the more dangerous and well known foodborne disease agent. (Note: *Staph* also contaminate cosmetics, medical supplies etc.) Food contamination by this microorganism is virtually always because of human contamination. When this bacteria is allowed to grow in foods, usually after human contamination and improper holding temperatures, it can produce a toxin. Tests that measure the presence of coagulase-positive *Staph* are used to indicate the likely presence of *Staph aureus*. Generally, finding even low counts of *Staph. aureus* in prepared foods might mean that irradiation should not be used since it will not destroy the toxin that can cause illness and could kill the spoilage organisms that might otherwise cause the food to be thrown out. A finding of *Staph aureus* in a food presented for irradiation requires a lot of information about the food, its processing and storage history before accepting it for irradiation.

*Vibrio* – Gram-negative and very fast growing even at cold temperatures. Three *Vibrio* species are food or water pathogens. *Vibrio cholerae* is water borne, but *V. parahaemolyticus* and *V. vulnificans* come from seafood. Foodborne illness from *Vibrio* can be very serious/deadly.

*Yersinia* – Gram-negative toxin producer. The toxin is heat-stable, meaning this bacteria has to be quickly killed before it can develop toxin. Some are pathogens and one, *Y. pestis* is famous for causing the Plague. Food sources of *Y. enterocolitica* are usually animals, especially pigs, and raw milk, but *Yersinia* have also been found in fresh water fish or fowl, seafood, vac-packed meats.

### **Common Foodborne Fungi - Quick Notes**

Fungi usually cause spoilage, but some can also cause illness either directly, or because they produce toxins.

*Aspergillus* (mold) – responsible for black rot of several fruit, spoilage of ham and bacon and oils. While some are commercially used to make fermented foods, some of these fungi produce powerful toxins. The toxins are not radiation sensitive.

*Botrytis* (mold) – grey or blue mold on fruit such as apples, pears, berries and citrus.

*Fusarium* (mold) – common soil contaminant causes brown rot of fruit, and rots of wheat and barley. An inhalant danger.

*Saccharomyces* (yeast). More commonly used commercially for fermentation of beer wine, Champagne etc. Some of them do spoil juice, juice concentrates, salad dressings and wine. When acting as spoilage organisms, they usually act in concert with *Lactobacillus* bacteria.

### **Common Foodborne Parasites – Quick Notes**

*Giardia* – Mostly waterborne. Giardia can infect food through contamination with animal feces or contaminated water. Hikers and campers who are tricked into thinking cool mountain streams contain pure water can be infected with this parasite.

*Cryptosporidium parvum* – Pet reptiles are a frequent source of this parasite, but human or animal feces used as fertilizer has resulted in numerous cases from contaminated berries. Healthy adults may not become sick but this parasite is a serious infection for AIDS patients and other immunocompromised persons. Even healthy individuals have been known to produce 17 liters of stools in one day with the diarrhea that this parasite causes!

*Toxoplasma gondii* – Pet cats are the most frequent source of this parasite, but it is also found in meat. Adults may contract the parasite with no symptoms (usually by eating undercooked meat), but it also causes severe illness/deaths of human fetuses.

*Trichinella spiralis* – Raw or undercooked pork is the usual source of this parasite, but hunters consuming inadequately cooked bear meat have also contracted the disease – Trichinosis. Trichinosis rarely kills its victims – but as the parasite burrows into their muscles, its victims probably wish it did!

**Also note there many other parasites associated with fish and seafood that are not covered here; most cause spoilage and economic damage.**

## **Foodborne Viruses – Quick Notes**

**Hepatitis A.** - This is a virus of family Picornaviridae, a piece of information that is important only because it tells us this virus is in the same family as polio, Echovirus and Coxsackie virus (and because there has been irradiation research done on the radiation resistance of these viruses). They are single stranded RNA genomes, (not DNA virus like Smallpox). Hep A is usually a human health concern when infected individuals contaminate food. It has also been found to infect shellfish, fruits and vegetables when contaminated by feces.

**Norwalk and related viruses** – Waterborne and foodborne, Norwalk is usually implicated in large outbreaks and often in social setting involving children or older adults (camps, day care, holiday cruises etc.). Since it is usually caused by human contamination of foods that are then served without further cooking, it does not seem that irradiation has a part to play in prevention of Norwalk. I'm including it because it is becoming better known and you might be asked about it.

(Note: There are many known viruses, and probably many more that have not been well studied that are not included here.)

## How to Use the D<sub>10</sub> Value Table

For our purposes, a D<sub>10</sub> value is the amount of radiation required to kill 90% (one log) of the microorganism.

However, killing 90% of them is usually not enough. Depending on the common bioburden, or the perceived need for treatment, or the claim (e.g. pasteurization), you may want to target three or five times the treatment. This is called a 3D<sub>10</sub> or a 5D<sub>10</sub> treatment. Sterilization is generally considered to occur if a 12D<sub>10</sub> treatment for *Clostridium botulinum* is achieved.

A table of D<sub>10</sub> values can be misleading. As with other research, published D<sub>10</sub> values are sometimes very reliable and sometimes less reliable. Therefore, the results could be considered a likely estimate, or if the research has been replicated by several researchers and where the effect of different methods has been examined, the D<sub>10</sub> value may be exact. Much of the D<sub>10</sub> research has been conducted with laboratory strains, instead of naturally occurring strains. Naturally occurring strains can sometimes be more adapted and difficult to kill. For this reason, a published D<sub>10</sub> value cannot always tell you the radiation dose to kill that microorganism even when all the conditions might be the same. When conditions are different (eg. temperature, food type, pH, salt or water content, atmosphere), the D<sub>10</sub> value could be quite different.

Therefore when assisting customers to prepare HACCP plans or to plan confirmatory research, use the table of D<sub>10</sub> values to target and bracket the likely minimum dose.

One of the problems with relying just on D<sub>10</sub> values when trying to understand what the microbiological effect is that most of the research does not report the results in D<sub>10</sub> values. Consequently, a table of D<sub>10</sub> values only gives us about one-third of the research that might have been done with that food and that microorganism.

Generally, selecting a correct dose for processing requires the following pre-requisite knowledge:

- The type and usual bioburden of the microorganisms present (or potentially present),
- The regulated tolerance level for the microorganism, or if there is no regulatory guidance, it would be helpful to know other risk assessment factors such as the group most likely affected by the pathogen (children? Pregnant women? etc), or the human-infective dose of the microorganism, if possible,
- The D<sub>10</sub> of the most resistant or target microorganism in the product to be treated, under the conditions of processing to be used (Refrigerated?, Frozen?, Heat treated?, Type of packaging?).

Gathering information on the potential microbial hazards of the product, such as D<sub>10</sub> values, and knowledge of the regulated tolerance levels or risk of the microorganism, is part of hazard analysis under HACCP.

**Summary Table of D<sub>10</sub> Values from Published Research  
(alphabetical by microorganism)**

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<b>Microorganism or infective agent</b>	<b>Food source and environmental conditions</b>	<b>D<sub>10</sub> (kGy)</b>	<b>Reference</b>
<i>Achromobacter xyloxydans</i>	Frozen shrimp	0.052	Charbonneau et al, 1986
<i>Acinetobacter calcoaceticus</i> (catalase +) (psychrotroph) (catalase -)	Frozen shrimp	0.084 0.106 0.108	Charbonneau et al, 1986
<i>Aeromonas hydrophila</i> (five isolates)	2°C blue fish or ground beef	0.14-0.19	Palumbo et al, 1986 in Thayer 1992
	15° blue fish or ground beef	0.22-0.34	
<i>Aeromonas hydrophila</i>	Frozen shrimp	0.038	Charbonneau et al, 1986
<i>Agrobacterium ranchobacter</i> (psychrotroph)	Frozen shrimp	0.067	Charbonneau et al, 1986
<i>Alcaligenes faecalis</i>	Frozen shrimp	0.066	Charbonneau et al, 1986
<i>Arcobacter butzleri</i> (ATCC 49616)	Ground pork, in vacuum, on ice	0.27±4.8%	Grass prawns (Penaeus monodon) -10±2°C
<i>Aspergillus sp</i> (fungi)	Mangoes and Bananas	0.25	Alabastro et al, 1978
<i>Aspergillus flavus</i> (var. columnaris, several strains) <i>petrikii</i> <i>flavus</i> <i>parasiticus</i>	Spices, (range gives first wet and then dry conditions)	0.26-0.60 0.32-0.65 0/30-0.57 0.37-0.56	Kume et al, 1989

<b>Microorganism or infective agent</b>	<b>Food source and environmental conditions</b>	<b>D<sub>10</sub> (kGy)</b>	<b>Reference</b>
<i>Bacillus spp</i> ( <i>non-pigment psychrotroph</i> ) ( <i>pigmented psychrotroph</i> )	Frozen shrimp	0.088 0.166	Charbonneau et al, 1986
<i>Bacillus cereus</i>	3-4°C, Cooked cauliflower and potatoes	0.12-0.28	Grant and Patterson, 1992
<i>Bacillus cereus</i> (stationary cells)	5°C, in vacuum or in air Mechanically deboned chicken	0.45±0.02	Thayer and Boyd, 1994
<i>Bacillus cereus</i> (endospores)	5°C Mechanically deboned chicken Beef round Pork loin Turkey breast Beef gravy	1.92+0.072 2.78+0.173 2.78+0.315 1.90+0.113 2.77+0.205	Thayer and Boyd, 1994
<i>Bacillus cereus</i> (endospores)	5°C, in vacuum or in air Beef or pork	2.78	Thayer and Boyd, 1994
<i>Bacillus cereus</i>	5°C in air and in vacuum Mechanically deboned chicken Logarithmic cells Stationary cells Endospores	0.18±0.008 0.43±0.016 2.56±0.21	Thayer and Boyd, 1994
<i>Bacillus coli</i>	Frozen liquid egg	0.258	Fengmei et al, 2000
Bacteria (unspecified, and method unspecified)	Ground black pepper Paprika Thyme Marjoram Spice mix for goulash Spice mix for Frankfurters Spice mix for RTE pork sausage Spice mix for boiling sausages Mustard powder Curry powder	1.9 1.7 1.8 1.8 1.7 2.1 1.5 1.6 1.6 1.3	Wetzel et al, 1985

<b>Microorganism or infective agent</b>	<b>Food source and environmental conditions</b>	<b>D<sub>10</sub> (kGy)</b>	<b>Reference</b>
Bacteria (aerobes)	Powdered red pepper - e-beam Powdered red pepper - gamma Powdered ginger – e-beam Powdered ginger - gamma	1.5 1.6-1.8 1.3-2.2 1.4-2.7	Lee et al, 2000
Bacteria ( <i>unspecified, Total Viable Plate Count method</i> )	Coriander Cumin Turmeric Chili	2.14 1.67 1.67 1.07	Alam et al, 1992
Bacteria ( <i>unspecified, Total Plate Count method</i> )	Turmeric Black Pepper Chili	1.44 1.53 1.25	Lokendra Singh et al, 1988
Bacteria ( <i>unspecified, Total Plate Count method</i> )	White pepper Paprika Nutmeg	1.77 1.88 2.14-2.38	Calenberg et al, 1998
Bacteria ( <i>mesophiles, Standard Plate Count method</i> )	Paprika	2.12-2.14	Nieto-Sandoval et al, 2000
Bacteria (aerobes)	Paprika Whole Black Pepper	1.44 1.29	Kiss and Farkas, 1981
<i>Byssochlamys fulva</i> (mold)(ascospores, three strains)	Sterilized apple juice, 25°C	1.2	Van der Reit and Van der Walt, 1985
<i>Campylobacter jejuni</i>	Ground beef, 3-5°C, 8-14% fat Ground beef, -17-15°C, 8-14% fat Gound beef, 3-5°C, 27-28% fat Ground beef, -17-15°C, 27-28% fat	0.17±.005 0.23±.017 0.19±.017 0.20±0.16	Clavero et al, 1993
<i>Campylobacter jejuni</i> (three strains)	Raw beef, 18-20°C	0.14-0.16	Tarkowski et al, 1984

<b>Microorganism or infective agent</b>	<b>Food source and environmental conditions</b>	<b>D<sub>10</sub> (kGy)</b>	<b>Reference</b>
<i>Campylobacter jejuni</i> NCTC 11351 <i>jejuni</i> FN1 <i>jejuni</i> FN2 <i>coli</i> FM3 <i>coli</i> NCTC 11366 <i>coli</i> FM4 <i>fetus</i> NCTC 10842 <i>lari</i> NCTC 11352	Minced chicken, sterilized and then inoculated, 4°C	0.19 0.16 0.13 0.25 0.14 0.12 0.14 0.14	Patterson, 1995
<i>Campylobacter jujuni</i>	Ground pork, in vacuum, on ice	0.19±4.2%	Collins et al, 1996
<i>Clostridia</i> (Sulfide-reducing type)	Paprika	3.84-4.65	Nieto-Sandoval et al, 2000
<i>Clostridium botulinum</i> , Types A, B	Cured bacon, in vacuum, in cans	2.20 – 2.39	Anellis et al 1965, reported in Thayer 1986
<i>Clostridium botulinum</i> , Types A, B	Cured ham, cooked, in vacuum, in cans, -30°C	2.6	Anellis et al 1977a, reported in Thayer 1986
<i>Clostridium botulinum</i> , Types A, B	Corned beef; pork sausage, in vacuum, -30°C ±10°C	1.9 – 2.14	Anellis et al 1962, reported in Thayer 1986
<i>Clostridium botulinum</i> , Types A, B	Cooked, in vacuum, in cans, -30+10°C beef chicken pork loin	3.4 3.5 3.6	Anellis et al 1969, 1977b, 1979, reported in Thayer 1986
<i>Clostridium botulinum</i> , Types A,B	Cooked, in vacuum, in cans Pork Chicken	3.64 3.58	Anellis et al, 1977b

<b>Microorganism or infective agent</b>	<b>Food source and environmental conditions</b>	<b>D<sub>10</sub> (kGy)</b>	<b>Reference</b>
<i>Clostridium perfringens</i> (vegetative cells)	3-4°C Cooked minced beef	0.34-0.58	Grant and Patterson, 1992
<i>Clostridium perfringens</i>	Intestines used as natural sausage casings Small fresh pork casings Large fresh pork casings	5.68 5.07	Trigo and Fraqueza, undated
** <i>Clostridium perfringens</i> (range of two substrate and plating methods)	Cooked Roast beef Gravy Cauliflower Roast potatoes Mashed potatoes	0.79-0.58 0.36-0.41 0.47-0.52 0.42-0.48 0.34	Grant and Patterson, 1992
<i>Clostridium perfringens</i>	Ground pork, approx 4°C, summary of two plating methods In air 25%CO <sub>2</sub> :75%N <sub>2</sub>	0.79-0.826 0.67-0.75	Grant and Patterson, 1991
<i>Clostridium sporogenes</i> (ATCC 7955)	Chicken breast, in vacuum, on ice D10 for irradiation alone D10 for irradiation when proceeded by 6,800 atm at 80°C for 20 min	4.1 2.0	Crawford et al, 1996
<i>Coliforms</i> (unspecified)	Turmeric Black pepper Chili	1.04 0.96 1.0	Lokendra Singh et al, 1988
<i>Coliforms</i> (unspecified)	Coriander Cumin Turmeric Chili	0.36 0.63 0.42 0.71	Alam et al, 1992
<i>Coliforms</i> (unspecified)	Paprika	3.15-3.25	Nieto-Sandoval et al, 2000
<i>Colletotrichum sp</i> (fungi)	Mangoes and bananas	0.54	Alabastro et al, 1978
<i>Coxsackie virus</i>	Water Media Cooked frozen ground beef	1.2 4.8 8.1	Jacobs, 1994

<b>Microorganism or infective agent</b>	<b>Food source and environmental conditions</b>	<b>D<sub>10</sub> (kGy)</b>	<b>Reference</b>
<i>Enterobacteriaceae</i>	Intestines used as natural sausage casings Small fresh pork casings Large fresh pork casings Beef dry casings	0.27 0.33 1.48	Trigo and Fraqueza, undated
<i>Enterobacteriaceae</i>	Paprika	2.66-3.02	Nieto-Sandoval et al, 2000
<i>E. coli</i>	Grass prawns ( <i>Penaeus monodon</i> ) -10±2°C	0.39	Hau et al, 1992
<i>E. coli</i> (two plating mediums, summary results)	Minced chicken, sterilized then inoculated, 4°C In air CO <sub>2</sub> In vacuum N <sub>2</sub>	0.35-0.38 0.28-0.29 0.26-0.27 0.23-0.25	Patterson, 1988
<i>E. coli</i>	Ground pork, approx 4°C, summary of two plating methods In air 25%CO <sub>2</sub> :75%N <sub>2</sub>	0.33-0.34	Grant and Patterson, 1991
<i>E. coli</i>	5°C Beef Lamb Pork Turkey breast Turkey leg	0.30 ±0.02 0.32±0.02 0.30±0.01 0.30±0.01 0.29±0.04	Thayer et al, 1995
<i>E. coli</i> 0157:H7	In vacuum, 5°C Finely ground lean beef Lean ground beef (2.6% fat) Mechanically separated chicken (21.3% fat)	0.28 0.27 0.27	Thayer and Boyd, 1993
<i>E. coli</i> 0157:H7 (three strains, non-acid adapted and acid adapted cells)	Pasteurized, clarified apple juice	0.12-0.31	Buchanan et al, 1998

<b>Microorganism or infective agent</b>	<b>Food source and environmental conditions</b>	<b>D<sub>10</sub> (kGy)</b>	<b>Reference</b>
<i>E. coli</i> 0157:H7	Ground beef, 3-5°C, 8-14% fat	0.24±.012	Clavero et al, 1993
	Ground beef, -17-15°C, 8-14% fat	0.30±.015	
	Gound beef, 3-5°C, 27-28% fat	0.25±.028	
	Ground beef, -17-15°C, 27-28% fat	0.30±0.23	
<i>Echovirus</i>	Water Media	1.4 5.7	Jacobs, 1994
<i>Hepatitis A virus</i>	Artificially inoculated fruits and vegetables, room temperature	2.7 – 3.0	Jacobs, 1994; Bidawid et al, 2000
<i>Hepatitis A virus</i>	Clams, Oysters (live, 4°C, in air), numerous assay methods, (summary results, paper should be reviewed before research is started)	2.02	Mallett et al, 1991
<i>Hepatitis A virus</i>	Lettuce ambient temp	2.72±0.05	Bidawid, et al, 2000
	Strawberries ambient temp	2.97±0.18	
<i>Lactobacillus</i>	Shrimp (On ice, likely 1±1°C)	0.59	Angel, 1986
<i>Lactobacillus spp</i>	Frozen shrimp	0.102	Charbonneau et al, 1986
<i>Lactobacillus spp</i>	Minced chicken, sterilized then inoculated, 4°C		Patterson, 1988
	In air	0.59	
	CO <sub>2</sub>	0.40	
	In vacuum	0.50	
<i>Listeria monocytogenes</i>	5°C		Thayer et al, 1995
	Beef	0.45±0.03	
	Lamb	0.47±0.04	
	Pork	0.48±0.02	
	Turkey breast	0.50±0.03	
Turkey leg	0.47±0.03		

<b>Microorganism or infective agent</b>	<b>Food source and environmental conditions</b>	<b>D<sub>10</sub> (kGy)</b>	<b>Reference</b>
<i>Listeria monocytogenes</i> (strain ATCC 35152)	Boneless chicken and minced lamb, in air, 0°C and 5°C	0.5	Kamat and Nair, 1995
<i>Listeria monocytogenes</i> (two strains, summary results)	Ground pork, approx 4°C, In air 25%CO <sub>2</sub> :75%N <sub>2</sub>	0.51-0.64 0.51-0.70	Grant and Patterson, 1991
<i>Listeria monocytogenes</i>	White crab meat 4°C	0.59	Chen et al, 1996
<i>Listeria monocytogenes</i>	Beef (tenderloin) 0° -5°C -20°C	0.45 0.77 1.21	Thayer and Boyd, 1995
<i>Listeria monocytogenes</i>	Chicken meat	0.43	Huntanen et al, 1989 in Thayer et al 1995
<i>Listeria monocytogenes</i>	Chicken meat, ground	0.41-0.55	Patterson, 1988 in Thayer et al, 1995
<i>Listeria monocytogenes</i>	Ground beef	0.51-0.61	Beuchat et al, 1993 in Thayer et al, 1995
<i>Listeria monocytogenes</i>	3-4°C Cooked ground beef and cooked cauliflower and potatoes	0.30-0.64	Grant and Patterson, 1992
<i>Listeria monocytogenes</i>	4 ±1°C beef bologna 3.5% soy protein beef bologna 1.75% soy protein beef bologna 0% soy protein	0.66 0.68 0.71	Sommers et al 2001
<i>Listeria monocytogenes</i>	Ground beef, 3-5°C, 8-14% fat Ground beef, -17-15°C, 8-14% fat Gound beef, 3-5°C, 27-28% fat Ground beef, -17-15°C, 27-28% fat	0.58+.063 0.61+.045 0.57+.039 0.57+.030	Monk et al, 1994

<b>Microorganism or infective agent</b>	<b>Food source and environmental conditions</b>	<b>D<sub>10</sub> (kGy)</b>	<b>Reference</b>
<i>Listeria monocytogenes</i>	4+1°C Beef frankfurters (product 1) Beef frankfurters (product 2) Mixed meat frank (product 1) Mixed meat frank (product 2) Poultry frank (product 1) Poultry frank (product 2) Poultry frank (product 3)	0.52±0.09 0.52±0.09 0.71±0.09 0.71±0.07 0.49±0.12 0.70±0.09 0.64±0.11	Sommers and Thayer, 2000
<i>Listeria monocytogenes</i> (ATC19111)	Sterilized chicken meat, on ice (summary of three dose rates)	0.36-0.38	Dion et al, 1994
<i>Listeria monocytogenes</i>	Shrimp, previously radiation sterilized and then artificially contaminated FDA-1A1 stationary phase 0°C FDA-1A1 stationary phase – 20°C ATCC 19115 stationary phase 0°C ATCC 19115 stationary phase –20°C	0.27-0.28 0.67-0.74 0.34-0.36 0.81-0.84	Brandao-Areal, et al, 1995
<i>Micrococcus spp</i>	Frozen shrimp	0.509	Charbonneau et al, 1986
<i>Mold (unspecified)</i>	Coriander Cumin Turmeric Chili	2.14 1.25 0.83 0.71	Alam et al, 1992
<i>Molds and yeasts (unspecified)</i>	Paprika	3.36-3.86	Nieto-Sandoval et al, 2000
<i>Moraxella spp</i> (MacConkey – psychrotroph) (MacConkey + psychrotroph)	Frozen shrimp	0.379 0.426	Charbonneau et al, 1986

<b>Microorganism or infective agent</b>	<b>Food source and environmental conditions</b>	<b>D<sub>10</sub> (kGy)</b>	<b>Reference</b>
<i>Moraxella phenylpyruvica</i>	Minced chicken, sterilized then inoculated, 4°C In air CO <sub>2</sub> In vacuum N <sub>2</sub>	0.85 0.67 0.62 0.88	Patterson, 1988
<i>Polio virus</i>	Water Media	1.1 5.4	Jacobs, 1994
<i>Polio virus</i>	Clams, Oysters (live, 4°C, in air), numerous assay methods, (summary results, paper should be reviewed before research is started)	3.1	Mallett et al, 1991
<i>Pseudomonas putrefaciens</i> (catalase –) <i>aeruginosa</i> (psychrotroph) <i>paucimobilis</i> spp catalase -	Frozen shrimp	0.028 0.036 0.037 0.091	Charbonneau et al, 1986
<i>Pseudomonas putida</i>	Minced chicken, sterilized then inoculated, 4°C In air CO <sub>2</sub> In vacuum N <sub>2</sub>	0.08 0.110 0.06 0.08	Patterson, 1988
<i>Rotavirus</i>	Clams, Oysters (live, 4°C, in air), numerous assay methods, (summary results, paper should be reviewed before research is started)	2.4	Mallett et al, 1991

<b>Microorganism or infective agent</b>	<b>Food source and environmental conditions</b>	<b>D<sub>10</sub> (kGy)</b>	<b>Reference</b>
<i>Salmonella spp</i>	5°C Beef Lamb Pork Turkey breast Turkey leg	0.70±0.04 0.67±0.04 0.51±0.03 0.71±0.04 0.71±0.04	Thayer et al, 1995
<i>Salmonella</i>	Ground beef, 3-5°C, 8-14% fat Ground beef, -17-15°C, 8-14% fat Gound beef, 3-5°C, 27-28% fat Ground beef, -17-15°C, 27-28% fat	0.62±.089 0.80±.054 0.66±.031 0.74±.057	Beuchat et al, 1993
<i>Salmonella spp</i>	5°C Bison Ostrich Alligator Caiman	0.55±0.03 0.50±0.01 0.54±0.02 0.53±0.04	Thayer et al, 1997
<i>Salmonella</i>	Egg powder	1.0	Narvaiz et al, 1992
<i>Salmonella anatum</i> <i>arizonae</i> <i>newport</i> <i>enteritidis</i> <i>enteritidis</i> <i>newport</i> <i>typhimurium</i> <i>typhimurium</i>	In air and in vacuum, 4°C Mechanically deboned chicken Meat	0.52±0.114 0.44±0.057 0.53±0.107 0.56±0.095 0.77±0.095 0.37±0.030 0.39±0.010 0.51±0.034	Thayer et al, 1990
<i>Salmonella enteritidis</i>	Orange juice (five formulations including calcium added)	0.35-0.37	Niemira, 2001
<i>Salmonella enteritidis</i>	Grass prawns ( <i>Penaeus monodon</i> ) -10±2°C	0.48	Hau, 1992
<i>Salmonella</i> (mixtures of <i>S. lille</i> , <i>S. enteritidis</i> and <i>S. typhimurium</i> )	Egg powder	0.8	Matic et al, 1990.

<b>Microorganism or infective agent</b>	<b>Food source and environmental conditions</b>	<b>D<sub>10</sub> (kGy)</b>	<b>Reference</b>
<i>Salmonella typhimurium</i> (strain SR-11) (strain RIA)	Sterilized chicken, 4°C	0.52 0.66	Previte et al, 1970
<i>Salmonella typhimurium</i> ; <i>Salmonella stanley</i>	18-20°C Raw ground beef	0.55 0.78	Tarkowski et al 1984 in Thayer et al, 1995
<i>Salmonella typhimurium</i> (two media methods, summary results)	Minced chicken, sterilized then inoculated, 4°C In air CO <sub>2</sub> N <sub>2</sub>	0.43-0.50 0.44-0.54 0.55-0.62	Patterson, 1988
<i>Salmonella typhimurium</i> (two strains, summary results)	Ground pork, approx 4°C, summary of two plating methods In air 25%CO <sub>2</sub> :75%N <sub>2</sub>	0.41-0.86 0.37-0.92	Grant and Patterson, 1991
<i>Salmonella typhimurium</i> (two strains)	In air, 10°C Pork	0.40 – 0.86	Patterson 1991 in Thayer et al, 1995
<i>Salmonella typhimurium</i>	Soft shell clams (probably room temperature)	0.58-0.64	Licciardello et al, 1989
<i>Salmonella typhimurium</i>	3-4°C Cooked minced beef	0.37-0.69	Grant and Patterson, 1992
<i>Salmonella typhimurium panama anatum Stanley</i>	Raw beef, 18-20°C	0.55 0.66 0.67 0.78	Tarkowski et al, 1984
<i>Salmonella typhimurium</i> (ATCC 14028)	Sterilized chicken meat, on ice (summary of three dose rates)	0.31-0.33	Dion et al, 1994

Microorganism or infective agent	Food source and environmental conditions	D <sub>10</sub> (kGy)	Reference
<i>Shigella flexneri</i>	Soft shell clams (probably room temperature)	0.30-0.35	Licciardello et al, 1989
<i>Staphylococcus spp</i>	Frozen shrimp	1.121	Charbonneau et al, 1986
<i>Staph. aureus</i>	5°C beef lamb pork turkey breast turkey leg mechanically deboned chicken	0.46±0.02 0.40±0.03 0.43±0.02 0.45±0.03 0.45±0.05 0.41±0.03	Thayer et al, 1995
<i>Staph. aureus</i>	Minced chicken, sterilized then inoculated, 4°C In air CO <sub>2</sub> In vacuum N <sub>2</sub>	0.41 0.41 0.39 0.37	Patterson, 1988
<i>Staph. aureus</i>	Grass prawns ( <i>Penaeus monodon</i> ) -10±2°C	0.029	Hau et al, 1992
<i>Staph. aureus</i>	White crab meat 4°C	0.16	Chen et al, 1986
<i>Staph. aureus</i>	5°C Bison Ostrich Alligator Caiman	0.40±0.02 0.34±0.02 0.36±0.01 0.38±0.01	Thayer et al, 1997
<i>Staph. aureus</i> (strain ATCC 13565)	Sterilized, mechanically deboned chicken, 0°C Stationary phase cells, combined results of in vacuum and in air Mid-log phase cells in vacuum	0.36±0.01 0.27±0.02	Thayer and Boyd, 1992
<i>Staph. aureus</i>	Various atmospheres Chicken mince	0.37-0.42	Patterson, 1988, in Thayer et al, 1995
<i>Staph. aureus</i>	Low fat beef High fat beef	0.58 0.30	Maxcy and Tiwari, 1973 in Thayer et al, 1995

<b>Microorganism or infective agent</b>	<b>Food source and environmental conditions</b>	<b>D<sub>10</sub> (kGy)</b>	<b>Reference</b>
<i>Staph. aureus</i>	Ground beef, 3-5°C, 8-14% fat Ground beef, -17-15°C, 8-14% fat Ground beef, 3-5°C, 27-28% fat Ground beef, -17-15°C, 27-28% fat	0.45±.035 0/45±.038 0.44±.022 0.43±0.47	Monk et al, 1994
<i>Staph. aureus</i>	Soft shell clams	0.80-0.89	Licciardello et al, 1989
<i>Streptococci spp</i>	Frozen shrimp	0.137	Charbonneau et al, 1986
<i>Streptococci</i> (faecal origin)	Intestines used as natural sausage casings Small fresh pork casings Large fresh pork casings Beef dry casings	1.25 2.24 5.53	Trigo and Fraqueza, undated
<i>Streptococcus faecalis</i>	Soft shell clams (probably room temperature)	0.97-1.10	Licciardello et al, 1989
<i>Streptococcus faecalis</i>	Minced chicken, sterilized then inoculated, 4°C In air CO <sub>2</sub> In vacuum N <sub>2</sub>	0.65 0.70 0.69 0.67	Patterson, 1988
<i>Vibrio spp</i>	White crab meat	<0.10	Chen et al, 1996
<i>Vibrio cholera</i>	Grass prawns ( <i>Penaeus monodon</i> ) -10±2°C	0.11	Hau et al, 1992

Microorganism or infective agent	Food source and environmental conditions	D <sub>10</sub> (kGy)	Reference
<i>Vibrio parahaemolyticus</i>	Frozen shrimp	0.10	Bandekar et al, 1987
<i>Vibrio vulnificans</i>	Shellstock oysters (oysters in shell) Virulent form Avirulent form	 0.062 0.037	Dixon et al, 1995
Yeasts and molds (unspecified)	Turmeric Black pepper Chili	1.23 1.14 1.21	Lokendra Singh et al, 1988
<i>Yersinia enterocolitica</i>	Beef, 25°C	0.11	El-Zawahry and Rowley, 1979 in Olson 1998
<i>Yersinia enterocolitica</i> (three strains)	Raw beef, 18-20°C	0.10-0.21	Tarkowski et al, 1984
<i>Yersinia enterocolitica</i> (NCIMB 11174)	Ground pork, approx 4°C, summary of two plating methods In air 25%CO <sub>2</sub> :75%N <sub>2</sub>	 0.16 0.17-0.18	Grant and Patterson, 1991
<i>Yersinia enterocolitica</i> (strain F5692)	Homogenate of raw meat and salami 0-5°C -40°C	 0.25 0.25	Kamat et al, 1997

**Notes:**

Where possible, D<sub>10</sub> values determined when the microorganism was in food were preferentially selected for reporting in this table. Usually D<sub>10</sub> measurements in media were omitted.

Although the correct abbreviation is S. for both *Staphylococcus* and/or *Salmonella*, for the purposes of this table I have used the abbreviation Staph. to prevent confusion.

Where 12D<sub>10</sub> values were reported by published authors, these were recalculated to give a D<sub>10</sub> value. According to Dr. Don Thayer, this gives us a fairly close D<sub>10</sub> value but since 12D<sub>10</sub> values have to include a shoulder for the kill curve for *C. botulinum*, it may be a bit high.

\*\* Grant and Patterson noted several differences in D<sub>10</sub> values of *C. perfringens* when different microbiological methods were used. The range in this table reflects both methods; the standard error was omitted since it was reflected in the range.

**Reference list – Food Microbiology Reference Tool  
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Aalabastro Estrella F., Pineda Alicia S., Pangan Africa C., and Mercedita J. del Valle. 1978. Irradiation of fresh Cavendish bananas (*Musa cavendishii*) and mangoes (*Mangifera indica* Linn. var. *carabao*): The microbiological aspect. Food Preservation by Irradiation, Proceedings of International Symposium organized by IAEA, FAO, WHO, Vienna. pp 283-303.

Alam M.K., Choudhury N., Chowdhury N.A., and Q.M. Youssouf. 1992. Decontamination of spices by gamma radiation. Letters in Applied Microbiology. Vol 14 pp 199-202.

Anellis A., Shattuck E., Morin M., Srisara B., Qvale S., Rowley D.B., and E.W. Ross. 1977. Cryogenic gamma irradiation of prototype pork and chicken and antagonistic effect between *Clostridium botulinum*. Applied and Environmental Microbiology. Vol 34 (6) pp 823-831.

Angel S., Juven B.J., Weinberg Z.G., Linder P., and E. Eisenberg. 1986. Effects of radurization and refrigerated storage on quality and shelf-life of freshwater prawns, *Macrobrachium rosenbergii*. Journal of Food Protection. Vol 49 (2) pp 142-145.

Bandekar J.R., Chander R., and D.P. Nerkar. 1987. Radiation control of *Vibrio parahaemolyticus* in shrimp. Journal of Food Safety. Vol 8 pp 83-88.

Bidawid S., Farber J.M., and S.A. Sattar. 2000. Inactivation of hepatitis A virus (HAV) in fruits and vegetables by gamma irradiation. International Journal of Food Microbiology. Vol 57 pp 91-97.

Brandao-Areal Henrique, Charbonneau Raymond, and Chantal Thibault. 1995. Effect of ionization on *Listeria monocytogenes* in contaminated shrimps. Science Aliments. Vol 15 (3) pp 261-272.

Buchanan R.L., Edelson S.G., Snipes K., and G. Boyd. 1998. Inactivation of *Escherichia coli* O157:H7 in apple juice by irradiation. Applied and Environmental Microbiology. Vol 64 (11) pp 4533-4535.

Charbonneau R., Dubois G., Micusan M., and M. Gagnon. 1986. Effects of gamma rays on the conservation of northern shrimps and on their bacterial flora. Sci. Aliments. Vol 6 (2) pp 245-256.

Chen Y.P., Andrews L.S., and R.M. Grodner. 1996. Sensory and microbial quality of irradiated crab meat products. Journal of Food Science. Vol 61 (6) pp 1239-1241.

Clavero M. Rocelle S., Monk J. David, Beuchat Larry R., Doyle Michael P., and Robert E. Brackett. 1994. Inactivation of *Escherichia coli* O157:H7, Salmonellae, and *Campylobacter jejuni* in raw ground beef by gamma irradiation. *Applied and Environmental Microbiology*. Vol 60 (6) pp 2069-2075.

Collins Clifford I., Murano Elsa A., and Irene V. Wesley. 1996. Survival of *Arobacter butzleri* and *Campylobacter jejuni* after irradiation treatment in vacuum-packaged ground pork. *Journal of Food Protection*. Vol 59 (11) pp 1164-1166.

Crawford Yolande J., Murano Elsa A., Olson Dennis G., and Kalpana Shenoy. 1996. Use of high hydrostatic pressure and irradiation to eliminate *Clostridium sporogenes* spores in chicken breast. *Journal of Food Protection*. Vol 59 (7) pp 711-715.

Dion Paule, Charbonneau Raymond, and Chantal Thibault. 1994. Effect of ionizing dose rate on the radioresistance of some food pathogenic bacteria. *Canadian Journal of Microbiology*. Vol 40 pp 369-374.

Dixon D.W., and G.E. Rodrick. 1998. Effect of gamma radiation on shellstock oysters. Extension of shelf-life and reduction in bacterial numbers with particular reference to *Vibrio vulnificus*. In: *Combination Processes for Food Irradiation*. Joint FAO/IAEA Division of Nuclear Techniques in Food and Agriculture. Vienna (Austria) IAEA pp 97-110.

Fengmei Li, Yongbao Gu, and Chen Dianhua. 2000. Study on radiation preservation of frozen egg liquid. *Radiation Physics and Chemistry*. Vol 57 pp 341-343.

Grant Irene R., and Margaret F. Patterson. 1991 Effect of irradiation and modified atmosphere packaged on the microbiological safety of minced pork stored under temperature abuse conditions. *International Journal of Food Science and Technology*. Vol 26 pp 521-533.

Grant I.R., and M. F. Patterson. 1992. Sensitivity of foodborne pathogens to irradiation in the components of a chilled ready meal. *Food Microbiology*. Vol 9 pp 95-103.

Hau Lung-Bin, Liew Ming-Hsin and Lea-Te Yeh. 1992. Preservation of grass prawns by ionizing radiation. *Journal of Food Protection*. Vol 55 (3) pp 198-202.

Jacobs G. Irradiation D10 values for various bacteria, viruses, molds and fungi. 1994. *Parenteral Drug Association*.

Jay, J. 1996. *Modern Food Microbiology*. Chapman and Hall.

Kamat A.S., Khare S., Doctor T., and P.M. Nair. 1997. Control of *Yersinia enterocolitica* in raw pork and pork products by gamma irradiation. *International Journal of Food Microbiology*. Vol 36 pp 69-76.

Kamat Annapurna S., and Madhusudanan P. Nair. 1995. Gamma Irradiation as a means to eliminate *Listeria monocytogenes* from frozen chicken meat. Journal of Science, Food and Agriculture. Vol 69 pp 415-422.

Kiss I., and J. Farkas. 1981. Combined effect of gamma irradiation and heat treatment on microflora of spices. In: Combination Processes in Food Irradiation. Joint IAEA and FAO International symposium, Sri Lanka pp 107-113.

Kume Tamikazu, Ito Hitoshi, Soedarman Harsono, and Isao Ishigaki. 1989. Radiosensitivity of toxigenic *Aspergillus* isolated from spices and destruction of aflatoxins by gamma-irradiation. Radiation, Physics and Chemistry. Vol 34 (6) pp 973-978.

Lee Jungeun, Kwon Oh-Jin, and Joong-Ho Kwon. 2000. Effects of Electron beam irradiation on microbiological and organoleptic qualities of powdered red pepper and ginger. Korean Journal of Food Science and Technology. Vol 32 (2) pp 380-386.

Licciardello Joseph J., D'entremont Daniel L., and Ronald C. Lundstrom. 1989. Radio-resistance of some bacterial pathogens in soft-shell clams (*Mya Arenaria*) and mussels (*Mytilus Edulis*) Journal of Food Protection. Vol 52 (6) pp 407-411.

Mallett John C., and Leon E. Beghian. 1991. Potential of irradiation technology for improved shellfish sanitation. Journal of Food Safety. Vol 11 pp 231-245.

Matic Stjepan, Mihokovic Vlado, Katusin-Razem Branka, and Dusan Razem. 1990. The Eradication of *Salmonella* in egg powder by gamma irradiation. Journal of Food Protection. Vol 53 (2) pp 111-114.

Monk J. David, Clavero MA. Rochelle S., Beuchat Larry R., Doyle Michael P. and Robert E. Brackett. 1994. Irradiation inactivation of *Listeria monocytogenes* and *Staphylococcus aureus* in low- and high fat, frozen and refrigerated ground beef. Journal of Food Protection. Vol 57 (11) pp 969-974.

Narvaiz P., Lescano G. and E. Kairiyama. 1992. Physicochemical and sensory analyses on egg powder irradiated to inactivate *Salmonella* and reduce microbial load. Journal of Food Safety. Vol 12 pp 263-282.

Neimira Brendan A., 2001. Citrus juice composition does not influence radiation sensitivity of *Salmonella* Enteritidis. Journal of Food Protection. Vol 64 (6) pp 869-872.

Nieto-Sandoval Jose M., Almela Luis, Fernandez-Lopez Jose A., and Jose A. Munoz. 2000. Effect of Electron beam irradiation on color and microbial bioburden of red paprika. Journal of Food Protection. Vol 63 (5) pp 633-637.

Olson Dennis G. 1998. Irradiation of food. Food Technology. Vol 52 (1) pp 57-63.

Patterson Margaret. 1988. Sensitivity of bacteria to irradiation on poultry meat under various atmospheres. *Letters in Applied Microbiology*. Vol 7 pp 55-58.

Patterson M.F. 1995. Sensitivity of *Campylobacter* spp. to irradiation in poultry meat. *Letters in Applied Microbiology*. Vol 20 pp 338-340.

Previte Joseph J., Chang Y. and H.M. El-Bisi. 1970. Effects of radiation pasteurization on *Samonella*. I. Parameters affecting survival and recovery from chicken. *Canadian Journal of Microbiology*. Vol 16 pp 465-471.

Serrano L.E. Murano E.A. Shenoy K., and D.G. Olson 1997. D values of *Salmonella enteritidis* isolates and quality attributes of shell eggs and liquid whole eggs treated with irradiation. *Poultry Science*. Vol 76 pp 202-205.

Singh Lokendra, Mohan M.A., Padwal Desai S.R., Sankaran R. and T.R. Sharma. 1988. The use of gamma irradiation for improving microbiological qualities of spices. *Journal of Food Science and Technology*. Vol 25 (6) pp 357-360.

Sommers C.H., and D.W. Thayer. 2000. Survival of surface-inoculated *Listeria monocytogenes* on commercially available frankfurters following gamma irradiation. *Journal of Food Safety*. Vol 20 pp 127-137.

Sommers C.H., Fan X., Niemira B.A. and A.P. Handel. 2001. Effect of ionizing radiation of beef bologna containing soy protein concentrate. *Journal of Food Safety*. Vol 21 pp 151-165.

Tarkowski J.A., Stoffer S.C.C., Beumer R.R., and E.H. Kampelmacher. 1984. Low dose gamma irradiation of raw meat. I. Bacteriological and sensory quality effects in artificially contaminated samples. *International Journal of Food Microbiology*. Vol 1 pp 13-23.

Thayer Donald W. 1992. Irradiation for control foodborne pathogens on meats and poultry. *Proceedings of International Conference of the Agricultural Research Institute Orlando, Florida* pp 21-44.

Thayer Donald W. 2000. Sources of variation and uncertainty in the estimation of radiation D-10 values for foodborne pathogens. *USDA Office of Risk Assessment and Cost-Benefit Analysis*. Vol 5 (4) pp 1-5.

Thayer D.W., and G. Boyd. 1992. Gamma ray processing to destroy *Staphylococcus aureus* in mechanically deboned chicken meat. *Journal of Food Science*. Vol 57 (4) pp 848-851.

Thayer Donald W., and Glenn Boyd. 1993. Elimination of *Escherichia coli* O157:H7 in meats by gamma irradiation. *Applied and Environmental Microbiology*. Vol 59 (4) pp 1030-1034.

Thayer Donald W., and Glenn Boyd. 1994. Control of Enterotoxic *Bacillus cereus* on poultry or red meats and in beef gravy by gamma irradiation. *Journal of Food Protection*. Vol 57 (9) pp 758-764.

Thayer Donald W., and Glenn Boyd. 1995. Radiation sensitivity of *Listeria monocytogenes* on beef as affected by temperature. *Journal of Food Science*. Vol 60 (2) pp 237-240.

Thayer Donald W., Boyd Glenn, Fox Jay B., and Leon Lakritz. 1997. Elimination by gamma irradiation of *Salmonella* spp. and strains of *Staphylococcus aureus* inoculated in bison, ostrich, alligator, and caiman meat. *Journal of Food Protection*. Vol 60 (7) pp 756-760.

Thayer Donald W., Boyd Glenn, Fox J.B., Lakritz L., and J.W. Hampson. 1995. Variations in radiation sensitivity of foodborne pathogens associated with the suspending meat. *Journal of Food Science*. Vol 60 (1) pp 63-67.

Thayer Donald W., Lachina R. Victor, Huhtanen Charles N. and Eugen Wierbicki. 1986. Use of irradiation to ensure the microbiological safety of processed meats. *Food Technology*. pp 159-162.

Thayer Donald W., Boyd Glenn, Muller Wayne S., Lipson Carol A., Hayne Walter C., and Steven H. Baer. 1990. Radiation resistance of *Salmonella*. *Journal of Industrial Microbiology*. Vol 5 pp 383-390.

Trigo M.J., and M.J. Fraqueza. 1998. Effect of gamma radiation on microbial population of natural casings. *Radiation Physics and Chemistry*. Vol 52 (1-6) pp 125-128.

Van Calenberg S., Vanhaelewyn Gauthier, Van Cleemput Oswald, Callens Freddy, Mondelaers Wim and Andre Huyghebaert. 1998. Comparison of the effect of X-ray and electron beam irradiation on some selected spices. *Academic Press*. Vol 31 (3) pp 252-258.

Van Der Riet W.B. and W.H. Van Der Walt. 1985. Effect of Ionizing Radiation on Ascospores of three strains of *Byssochlamys fulva* in apple juice. *Journal of Food Protection*. Vol 48 (12) pp 1016-1018.

Wetzel K., Huebner G., and M. Baer. 1985. Irradiation of onions, spices and enzyme solutions in the German Democratic Republic. *Proceedings from International Symposium on Food Irradiation Processing IAEA-SM 271/16* pp 35-45.